

CHAPTER 3.9.5

CYSTICERCOSIS*¹

SUMMARY

Definition and description of the disease: *Cysticercosis of farmed and wild animals is caused by the larval stages (metacestodes) of cestodes of the family Taeniidae (tapeworms), the adult stages of which occur in the intestine of humans, dogs or wild Canidae. Bovine cysticercosis (primarily in muscle) and porcine cysticercosis (primarily in muscle, the central nervous system [CNS] and the liver) are caused by the metacestodes (cysticerci) of the human cestodes Taenia saginata and T. solium, respectively. Cysticerci of T. solium also develop in the CNS and musculature of humans. Taenia asiatica is a less widespread cause of cysticercosis in pigs, with the cysts locating in the liver and viscera and the adult tapeworm occurring in humans. Cysticercosis and coenurosis of sheep and goats, and occasionally cattle, with the cysts occurring in the muscles, brain, liver or peritoneal cavity, are caused by T. ovis, T. multiceps and T. hydatigena, with the adult tapeworms occurring in the intestines of dogs and wild canids.*

Most adult and larval tapeworm infections cause little or no disease. Exceptions are severe, potentially fatal human neurocysticercosis (NCC) caused by T. solium, and occasionally neurocoenurosis caused by T. multiceps in humans. These parasites are also occasional causes of muscle or ocular signs in humans. 'Gid' caused by T. multiceps in ruminants can require surgery or slaughter of the animal. Acute T. multiceps coenurosis and T. hydatigena cysticercosis in sheep and goats are rare but may be fatal. Cysticercosis causes economic loss through condemnation of infected meat and offal.

Identification of the agent: *Adult Taenia tapeworms are dorsoventrally flattened, segmented and large, reaching from 20 to 50 cm (species in dogs) to several metres (species in humans). Anteriorly, the scolex (head) has four muscular suckers and may have a rostellum, often armed with two rows of hooks, the length and number of these being relatively characteristic of a species. A neck follows the scolex, and this is followed by immature and then by mature reproductive segments, and finally gravid segments filled with eggs. Segment structure, although unreliable, can aid in the identification of the species. Adult Taenia are recognised at post-mortem or by passage of segments or eggs in faeces. Taenia species cannot be differentiated by egg structure. Metacestodes consist of a fluid-filled bladder with one or more invaginated protoscoleces. These 'bladderworms' are each contained within a cyst wall at the parasite–host interface. This structure comprises the cysticercus or coenurus. Metacestodes are grossly visible at post mortem and meat inspection, but light infections are often missed. NCC can be diagnosed by imaging techniques.*

Immunological tests: *Adult human Taenia infections can be recognised by detection of Taenia coproantigen in faeces using an antigen-capture enzyme-linked immunosorbent assay (Ag-ELISA), but the test does not differentiate species. A commercial test is available for detection of circulating parasite-derived antigen in the serum of cattle or pigs or humans with T. saginata or T. solium cysticercosis. Use of species-specific DNA-based techniques remains experimental.*

Serological tests: *Tests for antibodies in serum are not used currently for the diagnosis of cysticercosis in animals except for epidemiological purposes. Tests are available for the serological diagnosis of NCC in humans.*

¹ Although certain diseases caused by *Taeniidae* are included in some individual species sections of the OIE List, this chapter covers several species and thus gives a broader description.

Requirements for vaccines: Excellent vaccine antigens have been identified for the metacestodes, but not for the adult stages of *T. ovis*, *T. multiceps*, *T. saginata* and *T. solium*. A *T. ovis* vaccine was registered in New Zealand, but is not commercially available. A *T. solium* vaccine is undergoing the steps towards registration and commercial availability. A combination of vaccination and oxfendazole (OFZ) treatment was highly effective in experimental control of natural transmission to pigs.

A. INTRODUCTION

The metacestodes (or larval cestodes) of *Taenia* spp. tapeworms are the cause of cysticercosis in various farmed and wild animals and in humans. Adult tapeworms are found in the small intestine of carnivore definitive hosts: humans, dogs, and wild canids. *Taenia saginata* of humans causes bovine cysticercosis, which occurs virtually world-wide, but particularly in Africa, Latin America, Caucasian and South/Central Asia and eastern Mediterranean countries. The infection occurs in many countries in Europe and sporadically in the United States of America (USA), Canada, Australia and New Zealand. *Taenia solium* of humans causes porcine cysticercosis and human neurocysticercosis (NCC). It is found principally in Mexico, Central and South America, sub-Saharan Africa, non-Islamic countries of Asia, including India and China (People's Rep. of) in regions with poor sanitation and free-ranging, scavenging pigs. The cysticerci of *T. asiatica* of humans in South-East Asia occur in the liver of pigs. Dogs and wild canids are the definitive hosts of metacestodes of sheep, goats and other ruminants, which occur throughout most of the world, although *T. multiceps* has disappeared from the USA, Australia and New Zealand. *Taenia ovis* occurs in the muscles of sheep, *T. multiceps* in the brain (occasionally in the muscles) of sheep, goats, sometimes other ruminants and rarely humans, and *T. hydatigena* is found in the peritoneal cavity and on the liver of ruminants and pigs. Diagnosis in animals usually is based on identification of the metacestode at meat inspection or necropsy. Adults in definitive hosts are acquired by the ingestion of viable metacestodes in meat and offal that has not been adequately cooked or frozen to kill the parasite.

Gravid segments are shed by the adult tapeworms. These release about half their eggs through the site of the break into the faecal mass. Approximately 50% of the segments of *T. saginata* and the canid taeniids then migrate spontaneously out of the anus to fall to the ground, the rest of the segments are passed with the faeces. The segments migrate shedding most of the remainder of their eggs on the soil and herbage or on the faeces, respectively. Eggs may be disseminated from faeces by physical means or transport hosts. Flies particularly ingest eggs and transport these eggs, so eggs are deposited at high intensity within 150 m of the faeces and at low intensity for 10 km (Lawson & Gemmell, 1990). *Taenia solium* segments, however, are often passed in chains and flies are unimportant in dissemination. Eggs are immediately infective when passed. Animals acquire infection from ingestion of food or water contaminated with sticky eggs, ingestion of segments or faeces containing eggs. It is possible that pigs also acquire *T. solium* by coprophagy of the faeces of pigs that have eaten segments. Humans may be infected with *T. solium* by eggs on vegetables, in water, etc., that have been contaminated by faeces, or food contaminated by dirty hands, by faeco-oral transmission or through retroperistalsis and hatching of eggs internally. Disease clusters where a human carrier exists. Routine diagnosis of taeniosis continues to be mainly based on the morphology of the adult tapeworm and the presence of eggs or segments in the faeces of infected definitive hosts.

Taenia spp. are classed in Risk Group 2 for human infection and should be handled with appropriate measures as described in Chapter 1.1.4 *Biosafety and biosecurity: Standard for managing biological risk in the veterinary laboratory and animal facilities*. Biocontainment measures should be determined by risk analysis as described in Chapter 1.1.4.

B. DIAGNOSTIC TECHNIQUES

1. Identification of the agent

1.1. *Taenia saginata* (the beef tapeworm)

The adult is large, 4–8 metres long and can survive many years, usually singly, in the small intestine of humans. The scolex (or head) has no rostellum or hooks. Useful morphological features are presented in Table 1 (Khalil *et al.*, 1994; Loos-Frank, 2000; Soulsby, 1982; Verster, 1969). Gravid segments have >14 uterine branches. They usually leave the host singly and many migrate spontaneously from the anus.

The eggs are typical 'taeniid' eggs that cannot be differentiated morphologically from other *Taenia* or *Echinococcus* spp. eggs. *Taeniid* eggs measure about 25–45 µm in diameter; contain an oncosphere (or hexacanth embryo) bearing three pairs of hooks; have a thick, brown, radially striated embryophore or 'shell' composed of blocks; and there is an outer, oval, membranous coat, the true egg shell, that is lost from faecal eggs.

Metacestodes (*Cysticercus bovis*) of *T. saginata* usually occur in the striated muscles of cattle (beef measles), but also buffalo, and various *Cervidae*. Viable cysts are oval, fluid-filled, about 0.5–1 × 0.5 cm, translucent and contain a single white scolex that is morphologically similar to the scolex of the future adult tapeworm. They are contained in a thin, host-produced fibrous capsule. Cysts occasionally are found in the liver, lung, kidney, fat and elsewhere.

1.2. *Taenia solium* (the pork tapeworm)

Taenia solium is typically smaller than *T. saginata* being 1–5 metres and seems to survive for a shorter period, from a few months to 1 year. The scolex has an armed rostellum bearing two rows of hooks. Gravid segments have <14 uterine branches and do not usually leave the host spontaneously, but passively in chains with the faeces.

Metacestodes (*C. cellulosae*) occur in the muscles and central nervous system (CNS) of pigs (pork measles), bear and dogs, and in the muscles, subcutaneous tissues, CNS and, rarely the eye, of humans. Cysts are grossly similar to those of *T. saginata*. They have a scolex bearing a rostellum and hooks similar to the adult. Occasionally, in the brain cisterns of humans, cysts can develop in the space available as racemose cysts up to 10 cm or more across that lack a scolex.

Table 1. Useful features for identification of scoleces and segments of *Taenia* spp.

Parasite species	Number of hooks	Length of hooks (µm)		No. testes	Layers of testes	Cirrus sac extends to longitudinal vessels	No. uterine branches	
		Large hooks	Small hooks					
<i>T. hydatigena</i>	28–36 (26–44)	191–218 (170–235)	118–143 (110–168)	600– 700	1	Yes	6–10 that re-divide	Lobes of ovary unequal in size. No vaginal sphincter. Testes extend to vitellarium, but not confluent behind.
<i>T. ovis</i>	30–34 (24–38)	170–191 (131–202)	111–127 (89–157)	350– 750	1	No	11–20 that re-divide	Lobes of ovary unequal in size. Well developed vaginal sphincter. Testes extend to posterior edge of ovary.
<i>T. multiceps</i>	22–30 (20–34)	157–177 (120–190)	98–136 (73–160)	284– 388	2	Yes	14–20 that re-divide	Lobes of ovary equal in size. Pad of muscle on anterior wall of vagina. Testes extend to vitellarium, but not confluent behind.
<i>T. saginata</i>	– without rostellum	–	–	765– 1200	1	No	14–32 that re-divide Ratio of uterine twigs to branches 2.3	Lobes of ovary unequal in size with small Well developed vaginal sphincter. Testes extend to vitellarium, but not confluent behind.

Parasite species	Number of hooks	Length of hooks (µm)		No. testes	Layers of testes	Cirrus sac extends to longitudinal vessels	No. uterine branches	
		Large hooks	Small hooks					
<i>T. solium</i>	22–36	139–200	93–159	375–575	1	Yes	7–14 that re-divide	Lobes of ovary unequal in size with small accessory lobe. No vaginal sphincter. Testes confluent behind vitellarium
<i>T. asiatica</i>	Vestigial hooks some with small rostellum	–	–	868–904		No	16–32 that re-divide Ratio of uterine twigs to branches 4.4	Ovary, vaginal sphincter and extent of testes as <i>T. saginata</i> . Posterior protuberances on some gravid segments

1.3. *Taenia asiatica* (Asian *Taenia*)

Closely related to but genetically distinguishable from *T. saginata*, the adult in humans has an ovary, vaginal sphincter muscle and cirrus sac like those of *T. saginata*, but *T. asiatica* has a small rostellum and posterior protuberances on some segments and 16–32 uterine buds with 57–99 uterine twigs on one side. Segments are passed singly and often spontaneously.

The metacestodes (*C. viscerotropica*) are small, about 2 mm, and have a rostellum and two rows of primitive hooks, those of the outer row being numerous and tiny. They occur mainly in the parenchyma and on the surface of the liver of domesticated and wild pigs; they may be found on the mesenteries and, rarely, are described in cattle, goats, and monkeys.

1.4. *Taenia ovis*

Adults in the intestine of dogs and wild canines reach 1–2 metres in length and have an armed rostellum; the number and size of hooks can aid differentiation of *Taenia* spp. (Table 1). Metacestodes (*C. ovis*) that occur in the musculature (skeletal and cardiac) of sheep and less commonly goats reach 0.5–1.0 × 0.5 cm. A similar parasite (*T. ovis krabbei*) occurs in wild canines and dogs and the muscles of reindeer and deer in northern areas.

1.5. *Taenia hydatigena*

Adults are up to 1 metre or more long, are found in the intestine of dogs and wild canids, and have an armed rostellum (Table 1). Metacestodes (*C. tenuicollis*) can be large, from 1 cm up to 6–7 cm, and the scolex has a long neck. They are found attached to the omentum, mesentery and occasionally protruding from the liver surface, particularly of sheep, but also of other domesticated and wild ruminants and pigs. A wolf and reindeer/deer cycle exists in northern latitudes, in which the metacestodes are found in the liver of the intermediate host; canids are definitive hosts.

1.6. *Taenia multiceps*

Adults, up to a metre long in the intestine of canids, have an armed rostellum (Table 1). The metacestodes (*Coenurus cerebralis*) are large, white fluid-filled cysts that may have up to several hundred scoleces invaginated on the wall in clusters. Coenuri grow to 5 cm or more in size in the brain of sheep, the brain and intermuscular tissues of goats, and also the brain of cattle, wild ruminants and occasionally humans. The cysts induce neurological signs that in sheep are called 'gid', 'sturdy', etc.

1.7. Diagnosis of adult parasites in humans or canine carnivores

All parasite or faecal material from humans with possible *T. solium* infections must be handled with suitable safety precautions to prevent accidental infection with the eggs. *Taenia multiceps* and *Echinococcus* spp. also infect humans and, as taeniid eggs in dogs cannot be differentiated to species or genus level, in areas where these are endemic, the same safety precautions apply. In addition to

Taenia spp., humans and canine carnivores may be infected by *Diphyllobothrium* and *Hymenolepis* spp., while six other cestode genera are recorded occasionally in humans. These are described by Lloyd (2011) and all can be differentiated from *Taenia* spp. by egg/proglottid morphology. Recently however, *T. taeniaeformis* with morphologically indistinguishable taeniid eggs was recorded in a child. In canids, *Echinococcus* spp. eggs cannot be distinguished from *Taenia* spp. eggs, but the presence of the former can be determined by tapeworm size and *Echinococcus* -specific antigen-capture enzyme-linked immunosorbent assay (Ag-ELISA) (Allan *et al.*, 1992). Other worms in canids, *Dipylidium*, *Diplopylidium*, *Mesocestoides* and *Diphyllobothrium* spp., have morphologically distinct eggs and proglottids (Lloyd, 2011; Soulsby, 1982).

Adult cestodes can be expelled from humans using an anthelmintic followed by a saline purgative and are identified on the basis of scolex and proglottid morphology. A self-detection tool was used in Mexico (Flisser *et al.*, 2011); medical staff in health centres are supplied with preserved tapeworm segments in a bottle and a manual of questions to ask patients to try to identify carriers. In animals, arecoline purgation has been useful; again, the recovered tapeworms are identified morphologically. Arecoline is no longer available as an anthelmintic, but can be obtained from chemical supply companies. As it has side-effects, old, infirm and pregnant animals should be excluded from treatment. A dose of 4 mg/kg should result in purgation in under 30 minutes, provided food has been withheld for several hours (i.e. administer to dogs with empty stomachs). Walking and abdominal massage of recalcitrant cases or enema for constipated dogs may avoid the use of a second dose (2 mg/kg), which should be given only sparingly. Fortunately, arecoline purgation is being replaced rapidly by the copro-antigen ELISA for *Echinococcus* spp. and perhaps in the future this will also be the case for *Taenia* spp. Tapeworms can be recovered after anthelmintic treatment, and require appropriate disposal.

Verster (1969) and Loos-Frank (2000) have given descriptions of parasitic diagnosis of all the *Taenia* spp. of humans and animals, their hosts and geographical distributions. Keys for identification are given by Khalil *et al.* (1994). Loos-Frank (2000) gives methods for mounting, embedding, sectioning and staining the proglottids. Worms, after relaxation in water, can be stained directly, although small worms should be fixed in ethanol for a few minutes. Alternatively, worms can be fixed and stored in 70% ethanol containing 10% lactic acid, the scolex and worm being stored separately. The rostellum, hooks and suckers of scoleces or protoscoleces should be cut off and mounted *en face* in Berlese's fluid (made by dissolving 15 g gum arabic in 20 ml distilled water and adding 10 ml glucose syrup and 5 ml acetic acid, the whole then being saturated with chloral hydrate, up to 100 g). The stain is lactic acid carmine: 0.3 g carmine is dissolved at boiling point in 42 ml lactic acid and 58 ml distilled water, 5 ml of 5% iron chloride solution ($\text{FeCl}_2 \cdot 4\text{H}_2\text{O}$) is added after cooling and can be used again to refresh older solutions. Specimens are allowed to sink in the stain in a vial and are left in the stain for some more minutes to allow the stain to penetrate. Specimens are then washed in 1-day-old tap water until blue in colour. They are then fixed in 50–70% ethanol and dehydrated under the slight pressure of plastic foil keeping the segments flat. Salicylic acid methyl ester is used as clearant.

When segments break from the end of the worm, some eggs are expelled in the intestine and can be found in the faeces. Approximately 50% of the segments of *T. saginata*, *T. asiatica* and the dog *Taenia* spp. may migrate spontaneously from the anus and this is likely to be noticed (>95% in the case of *T. saginata*). When the segments migrate, the sticky eggs are deposited in the perianal area and might be detected by application and examination of sticky tape. These signs are far less likely for *T. solium*. Segments of all three may be found on the faeces, but are passed intermittently. Eggs are voided into the faecal mass but the migrating segments void about half their eggs in trails on the surface of faeces (to be eaten by flies) and surrounding area and on the ground after falling from the anus. Even if a segment has shed all its eggs, it can be identified as a cestode by the many concentric calcareous corpuscles contained within its tissues. Faeces, after mixing to reduce aggregation, can be examined for eggs. Various techniques are used throughout the world and include ethyl acetate extraction and flotation. For the latter, NaNO_3 or Sheather's sugar solution (500 g sugar, 6.6 ml phenol, 360 ml water), with their higher specific gravities, are superior to saturated NaCl as flotation media for taeniid eggs. Flotation can be carried out in commercially marketed qualitative or quantitative flotation chambers or by centrifugal flotation that includes a modified Wisconsin technique (faeces, diluted in water, are sieved and centrifuged, the pellet is resuspended in sugar or Sheather's solution and centrifuged at 300 g for 4 minutes). Eggs adhering to the cover-slip can then be detected. Faecal egg examination will be less sensitive for *T. solium* than the other species. Species cannot be determined by egg morphology. Cheesbrough (2005; 2006) reports that *T. saginata* eggs can be differentiated from *T. solium* on staining with Ziehl-Neelsen as used for acid-fast bacilli: the striated embryophore of *T. saginata* is acid fast (stains red), that of *T. solium* is not acid fast. DNA probes, the polymerase chain reaction (PCR) and PCR restriction fragment length polymorphism (RFLP), have proved useful for differentiation though largely used experimentally to differentiate faecal eggs of *T. solium*, *T. saginata* and *T. asiatica* (Gasser & Chilton, 1995; Gonzalez *et al.*, 2004). While equally applicable to differentiation in dogs, the same examinations have not been done for *Taenia* spp.

An Ag-ELISA to detect *Taenia* coproantigen is available from Cestode Diagnostics, University of Salford² and can be developed independently if laboratory facilities are available (Allan *et al.*, 1992). This Ag-ELISA was developed experimentally by Allan *et al.* (1992) to detect coproantigen in dogs, and so, with appropriate controls, could be used to detect *Taenia* infection in this species. The technique, however, is only *Taenia*-genus specific. The test is a solid-phase, microwell assay with wells coated with polyclonal, rabbit anti-*Taenia*-specific antibody (TSA). The following is the basic technique.

1.7.1. Test procedure

- i) Faecal supernatants are recovered from fresh, frozen or formalinised (5% formalin at 4°C) faecal samples. The sample is vigorously shaken forming a slurry in an equal weight/volume of 0.15 M phosphate-buffered saline (PBS) containing 0.3% Tween 30. The supernatant is recovered by centrifugation at 2000 **g** for 30 minutes.
- ii) A soluble aqueous extract of non-gravid proglottids from *Taenia* are obtained following emulsification in PBS and centrifugation.
- iii) Hyperimmune rabbit antiserum is produced against the soluble proglottid extract and the IgG fraction is isolated by passage into and elution of the bound IgG from Protein A-Sepharose CL 4B. Some of the IgG fraction is conjugated to peroxidase type VI. The sera are stored in small quantities frozen at –20°C. Sera may need to be absorbed by packed normal dog faeces in a ratio 2/1 with mixing for 1 hour and recovered by centrifugation.
- iv) Flat-bottomed microtitre plates are coated with rabbit anti-*Taenia* IgG (protein content 5–25 µg/ml determined by UV spectrophotometry) using 100 µl/well, the antisera are diluted in 0.05 M NaHCO₃/Na₂CO₃ buffer, pH 9.6. Plates are incubated overnight at 4°C, the wells are washed three times with PBS/0.1% Tween, blocked with PBS/0.3% Tween for 1 hour and washed again. 100 µl of faecal supernatant containing 50% fetal calf serum is added and the plates are incubated for 1 hour and then washed three times. 100 µl of the peroxidase-conjugated anti-*Taenia* IgG (diluted 1/100 or 1/200) is added and the plates are incubated for 1 hour before washing three times. Substrate solution (100 µl of 5-aminosalicylic acid and 0.005% H₂O₂ in 0.1 M phosphate buffer containing 1 mM Na₂EDTA [ethylene diamine tetra-acetic acid] at pH 6.0) is added for 25 minutes and the result is read at 450 nm. Cut-off values are the mean plus 3 standard deviations of values for normal dog faeces.

1.8. Diagnosis of metacestodes

Taenia solium or *T. saginata* metacestodes might be palpable in the tongue but, both in the living animal and on post-mortem examination or meat inspection, tongue palpation is of diagnostic value only in pigs or cattle heavily infected with metacestodes; these will also be difficult to differentiate from large sarcocysts.

1.8.1. Meat inspection – the main diagnostic procedure

Metacestodes are visible first as very small, about 1 mm, cysts, but detection of these requires thin slicing of tissues in the laboratory. Many young cysts are surrounded by a layer or capsule of inflammatory cells (mononuclear cells and eosinophils being prominent histologically). The parasites' abilities to evade the immune response mean that later in infection, as the cyst matures, few inflammatory cells are present in its vicinity and the cysticercus in its intermuscular location is surrounded by a delicate fibrous tissue capsule.

In theory, cysts can be visualised or felt in tissues such as the tongue of heavily infected animals as early as 2 weeks after infection. Cysts are readily visible by 6 weeks and, when mature, are usually oval, about 10 × 5 mm or larger, with a delicate, fairly translucent, white parasite membrane and host capsule. Pale fluid within the cyst and the scolex, visible as a white dot within the cyst, usually invaginates midway along the long axis of the cyst.

At meat inspection many of the cysts detected, often as many as 85–100%, are dead. The rate at which cysts age and die and so degenerate varies with the parasite species and the tissue within which the cyst is embedded and possibly with host age at infection. In general, cysts tend to die more rapidly in the muscular predilection sites, such as the heart. The preferential distribution of parasites in these areas may be because of greater blood circulation to these

2 Information on availability of the Ag-ELISA for the detection of *Taenia* coproantigen for epidemiological studies or potential use can be obtained from Professor P.S. Craig, OIE Reference Expert on Echinococcosis (see Table given in Part 4 of this *Terrestrial Manual*)

muscles. Conversely, the higher rate of activity in these muscles may damage the parasites, allowing leakage of fluid and perhaps disrupting the parasites' abilities to evade the immune response. Cysts at different stages of viability and degeneration can be found in the same host. Death in skeletal muscles may occur within 2 months of infection of adult cattle with *T. saginata*, but cysts may remain viable for several years. Cysts of *T. hydatigena* in the peritoneal cavity of sheep and those of *T. solium* in pigs also have been described as surviving for long periods. *Taenia solium* cysts survive for many years in the brain of humans, and frequently symptoms begin only as the cyst begins to degenerate.

Degenerating cysts vary in appearance. Marked infiltration of eosinophils, macrophages, lymphocytes and collagen deposition thickens the capsule, which becomes opaque, but initially the cyst within remains apparently normal. The fluid gradually becomes colloid and inflammatory cells infiltrate. The cyst cavity becomes filled with greenish (eosinophilic) and then yellow, caseous material and is very unaesthetic, usually being larger in size and certainly more obvious in meat than the original viable cyst. Later the cyst may calcify. Where very young (without a scolex) or degenerate cysts need to be differentiated from other lesions, compression of the cyst, smears of the caseous contents and histological examination of haematoxylin and eosin (H&E) stained sections are used. Microscopic examination may reveal the calcareous corpuscles (concentric concretions of salts that are around 5–10 µm in size). These indicate a cestode origin of the tissue. The presence of hooks and their length together with knowledge of the host and tissue may aid in identification of cestode species. Experimentally, immunohistochemical staining has differentiated *T. saginata* cysts from non-*Taenia* structures. PCR would be useful, but is experimental, in the finding of a new cestode in a host species or geographical area from which, historically, the parasite was absent. In one study PCR identified only 50% of degenerate presumed *T. saginata* cysts (Abuseir *et al.*, 2006), whereas Eichenberger *et al.* (2011) using different primers identified 80% of calcified lesions.

After treatment of *T. saginata* and *T. solium* in cattle and pigs with drugs such as albendazole and oxfendazole (OFZ), the cysts may lose their fluid and collapse. The resultant lesion is smaller than lesions observed following natural death but can take 3–6 months to resolve. However, cysts that died before treatment of the animal will remain large and visible.

Meat inspection procedures vary with the parasite, i.e. zoonotic or not, and the host involved, the tissue involved, and the regulations within a country. Examinations tend to be more extensive with the zoonotic infections *T. saginata* and *T. solium*.

In general, meat inspection procedures consist of:

- i) Visual inspection of the carcass, its cut surfaces and the organs within it. This may reveal *T. saginata*, *T. solium* and *T. ovis* in the muscles, *T. hydatigena* on the liver, mesenteries or omentum, or *T. multiceps* in the brain.
- ii) The external and internal masseters and the pterygoid muscles are each examined and one or two incisions made into each, the cuts being parallel to the bone and right through the muscle.
- iii) The freed tongue is examined visually and palpated, particularly for *T. solium*.
- iv) The pericardium and heart are examined visually. The heart usually is incised once lengthwise through the left ventricle and interventricular septum so exposing the interior and cut surfaces for examination. Incisions may go from the base to the apex and regulations also may require additional, perhaps four, deep incisions into the left ventricle. Alternately, the heart may be examined externally and then internally after cutting through the interventricular septum and eversion.
- v) The muscles of the diaphragm, after removal of the peritoneum, are examined visually and may be incised.
- vi) The oesophagus is examined visually.
- vii) In some countries, the triceps brachii muscle of cattle is incised deeply some 5 cm above the elbow. Additional cuts into it may be made. The gracilis muscle also may be incised parallel to the pubic symphysis. These cuts are usually undertaken for *T. solium* in pigs. Such incisions into the legs are made, particularly in African countries as it is suspected that more parasites lodge in these muscles in working or range animals walking long distances because of the exercise and consequent increased blood flow to these muscles. Other countries may also require such incisions into the legs. However, as this devalues the meat, such incisions are made most commonly once one or more cysts have been found at the predilection sites so as to determine the extent of the infection.

Overall, the initial incision into any tissue is the most important, but additional incisions may be required either by the regulations or if cysts are found on the initial incision(s). Eichenberger *et al.* (2013) reported an increase in sensitivity by multiple incisions. Details on meat inspection are supplied by Herenda (2000).

Additional or fewer procedures may be required for specific parasites and the judgements on the carcass, viscera, offal and blood will vary dependent on *Taenia* spp. and regulations within a country. Judgement on infected carcasses will fall into three main categories: i) approve for human consumption; ii) partially condemn and pass the remainder of the carcass, but in the case of the zoonoses, *T. saginata* and *T. solium*, the carcass, meat and viscera must be treated; and iii) totally condemn heavily infected carcasses or emaciated diseased ones.

1.8.2. Meat inspection – species differentiation

i) *Taenia saginata*

a) Predilection site

Calves under 6 weeks or <32 kg are not usually examined. Predilection sites are the heart, tongue, masseters and diaphragm, presumably because they receive the greatest blood. Nonetheless, cysts may be found in any muscle of the body. If one carcass in a lot is found to be infected, all carcasses from the same lot can be held until laboratory confirmation is obtained. If *T. saginata* infection is confirmed, additional incisions are usually made in the carcasses in the lot; all suspicious lesions found in the rest of the lot are considered to be *T. saginata* without laboratory confirmation. Lesions of *T. saginata* may need to be differentiated from *Sarcocystis* sarcocysts and other lesions. In PCR studies in Germany, Switzerland and New Zealand up to 20% of viable, presumed *T. saginata* cysts could not be positively identified, which suggests there may be an unidentified cestode infecting cattle (Abuseir *et al.*, 2006).

b) Judgement

If a carcass is considered to be heavily infected then the carcass, meat, offal and blood all are condemned. The description of a heavy infection varies, but generally it is the detection of cysts at two of the predilection sites plus two sites in the legs. In the case of a lesser infection, the infected parts and surrounding tissues are removed and condemned. Even a single dead cyst requires that the carcass and edible viscera must then be treated and this is justifiable as about 10% of lightly infected carcasses were found on dissection to have both dead and viable parasites within them. Treatment varies with country and facilities available and includes: i) freezing at lower than -10°C for >10 or 14 days, or lower than -7°C for 21 days; ii) boxes of boned meat are frozen at less than -10°C for >20 days; iii) heated to above 60°C throughout; iv) steamed at moderate pressure (0.49 kg/cm^2); v) heated at $95\text{--}100^{\circ}\text{C}$ for 30 minutes; or vi) pickled in salt solution for 21 days at $8\text{--}12^{\circ}\text{C}$. Blast freezing a 30 kg box generally is reported to require 2 × 24-hour cycles at -30.9°C followed by 72 hours cold storage at -23.3°C for death of scoleces. Often no treated meat can be exported although in some countries it can be exported in canned form.

ii) *Taenia solium*

a) Predilection site

The predilection sites are as for *T. saginata* although there are reports of higher prevalence in shoulder and thigh. Commonly one or more cuts are required 2.5 cm above the elbow joint. This is said to detect some 13% of infected carcasses that would otherwise have been missed.

b) Judgement

In some countries, any lightly or heavily infected pigs and their viscera and blood are condemned. In areas where infection is common, lightly infected carcasses can be passed for cooking and pickling and occasionally freezing.

iii) *Taenia asiatica*

a) Predilection site

The small size means detection of cysts is difficult except in heavy infection.

- b) Judgement
Condemnation of the liver.

- iv) *Taenia hydatigena*

- a) Predilection site

The parasite migrating in the liver leaves haemorrhagic tracks that then become green/brown with inflammation and later white due to fibrosis. For the records, these must be differentiated from those of liver flukes, if possible, by identification of the cysticerci or adult flukes. White spot from *Ascaris* infection is differentiated as the lesions appear as pale to white, small, isolated foci. Some cysts remain trapped below the liver capsule. These usually are small and degenerate early and then calcify into cauliflower-like lesions. Those that are retained at the liver surface are usually superficial and subserosal, while most of an *Echinococcus granulosus* hydatid cyst is deeper in the parenchyma. *Taenia hydatigena* cysts usually mature in the omental or mesenteric fat. If viable, the *T. hydatigena* cyst has a long-necked single scolex in virtually translucent cyst fluid. Fertile *Echinococcus* hydatid cysts have thicker walls and may contain many brood capsules containing protoscoleces; these appear as a sandy, whitish deposit within the cysts. Differentiation can be important in the implementation and monitoring of hydatid disease control measures so histology may be required. H&E-stained sections will reveal the laminated membrane of very young hydatid cysts as indicated by Lloyd *et al.* (1991). Its presence or absence can be confirmed by periodic acid–Schiff staining when the highly glycosylated proteins in the laminated membrane stain red. *Taenia hydatigena* lesions in cattle and pigs can be similar to tuberculosis. However, the portal and mesenteric lymph nodes are not involved, the contents of parasite cysts are more easily shelled-out and remainders of hooks and calcareous corpuscles may be seen or Ziehl–Neelsen staining may reveal bacteria.

- b) Judgement

Usually only a few cysts or tracks are present and these can be trimmed. Heavily infected livers and omentum are condemned. Rarely, acute infections are seen with large numbers of migrating parasites producing traumatic hepatitis, ascites, oedema, etc., and would result in secondary condemnation of the carcass.

- v) *Taenia multiceps*

- a) Predilection site

The parasites have a predilection site for the brain and spinal cord. Early migrating parasites can cause reddish haemorrhagic and later grey purulent tracks in the brain, and in heavy infections, the sheep may have a meningoencephalitis. Clinical signs caused by the mature cyst relate to pressure atrophy of adjacent nervous tissue and vary according to location in the brain. There may be impaired vision or locomotion if cysts are in the cerebral hemispheres and the sheep gradually may be unable to feed and will become emaciated. Cerebellar cysts may precipitate more acute and severe signs of ataxia or opisthotonus. In heavy infections, parasites migrate and begin development in other tissues, but they die early. These produce small lesions, 1 mm or so in size, that first contain an encapsulated cyst, then eosinophilic, caseous material that later may calcify.

- b) Judgement

Initially only the head is condemned or occasional cysts in intermuscular or subcutaneous sites are trimmed. With chronic infection, the animal may have been unable to feed, resulting in condemnation because of emaciation, etc.

- vi) *Taenia ovis*

- a) Predilection site

The predilection sites are as for *T. saginata*. Cysts may be confused with large *Sarcocystis gigantea* sarcocysts.

b) Judgement

Commonly detection of up to 2–5 cysts results in trimming and the carcass is passed. This does not prevent the unaesthetic presence of live or degenerate parasites in other tissues. Ultrasound and X-rays are being tested for detection of these. Some authorities may require that the meat be boned, trimmed and frozen or cooked. In heavy infections the carcass is condemned.

In general, meat inspection procedures detect only about 15–50% of the animals that are actually infected. Light infections are easily missed on palpation and meat inspection – in one study involving *T. saginata*, 78% of carcasses infected with >20 cysts were detected compared with those detected following dissection and slicing, while only 31% of those with fewer cysts were detected (Walther & Koske, 1980). Meat inspection efficacy will vary with the number and location of incisions (and the skill and experience of the inspector). For example, in Zimbabwe, 58% of cattle were positive in the head only, 20% in the shoulder only and 8% in the heart only, although overall 81% were found to be infected if all three organs were included. In Kenya Walther & Koske (1980) also found that the predilection sites were not necessarily infected in 57% of the cattle found positive on dissection. They also confirmed the importance of the shoulder incisions in detection of infection in African countries as 20% of the cattle found to be infected were positive in the shoulder only. Animals infected as very young calves may have few or no cysts in tongue and masseters. Wanzala *et al.* (2003), also in Kenya, described the insensitivity of meat inspection in detecting cysticerci: only 50% of naturally or artificially infected cattle were identified. Their observations indicated that a number of viable cysticerci may be missed.

In humans, the most common presenting sign in *T. solium* NCC is seizures followed by headache, but a range of signs, such as vomiting, psychoses, etc., are seen depending on the number, location and viability or level of degeneration of the cysticerci (viable, transitional dying, calcification) (Carpio *et al.*, 1994; Flisser *et al.* 2011). In humans, clinical evaluation and either computerised tomography (CT) scan or magnetic resonance imaging (MRI) detect the exact parenchymal and extraparenchymal locations and viability of *T. solium* and *T. multiceps* metacestodes and can follow the progression of the lesion. These remain the most efficient means of diagnosis, but access to imaging facilities may not be available in endemic areas. Calcified cysts in tissues are detected by radiography. The site of *T. multiceps* cysts in sheep brain might be identified by clinical signs presented and, possibly, softening of the skull overlying the coenurus.

1.9. Detection of circulating antigens

The development of an automated sensitive and specific diagnostic test would greatly reduce the costs of damage to the carcass and also the costs of labour at meat inspection. Sensitivity of serological tests for animals has not reached the stage where commercialisation for individual diagnosis or large-scale detection of infected carcasses in slaughter houses is possible. All assays tested – Ag-ELISA, antibody ELISA, enzyme-linked immunoelectro transfer blot (EITB) and tongue inspection – show low sensitivity in rural pigs infected naturally with low levels of *T. solium* (Dorny *et al.*, 2005; Sciuotto *et al.*, 1998). This finding is also true for *T. saginata* infections in cattle (Van Kerckhoven *et al.*, 1998). For example, only a small percentage (13–22%) of cattle carrying fewer than 30–50 viable cysticerci is detected by Ag-ELISA. Nonetheless, Ag-ELISAs do have a use in field-based epidemiological studies for indicating transmission. The detection of viable infections in cattle or pigs could indicate point sources of infection, season of transmission and age of animals at risk. The development of more sensitive and specific assays with recombinant antigens for diagnosis of NCC should improve immunodiagnosis of *T. solium* in pigs.

2. Serological tests for antibodies

Tests for circulating antibodies have little role in animals, except for epidemiological studies. A number of EITB and ELISAs for antibodies to *T. solium* in humans are now widely available. These were reviewed by Rodriguez *et al.* (2012) with comparisons of sensitivity and specificity.

C. REQUIREMENTS FOR VACCINES

No internationally accepted standard for the production of vaccines is available. Nonetheless, considerable information is available in the scientific literature immunogenic molecules, their recombinant technology and extraction, and their efficacy experimentally and in limited field trials. Such approaches offer the future prospect of technically effective vaccines provided that the cost–benefit analyses are favourable.

REFERENCES

- ABUSEIR S., EPE C., SCHNIEDER T., KLEIN G. & KÜHNE M. (2006). Visual diagnosis of *Taenia saginata* cysticercosis during meat inspection: is it unequivocal? *Parasitol. Res.*, **99**, 405–409.
- ALLAN J.C., CRAIG P.S., GARCIA NOVAL J., MENCOS F., LIU D., WANG Y., WEN H., ZHOU P., STRINGER R., ROGAN M. & ZEYHLE E. (1992). Coproantigen detection for immunodiagnosis of echinococcosis and taeniasis in dogs and humans. *Parasitology*, **104**, 347–355.
- CARPIO A., PLACENCIA M., SANTILLAN F. & ESCOBAR A. (1994). A proposal for classification of neurocysticercosis. *Can. J. Neurol. Sci.*, **21**, 43–47.
- CHEESBROUGH M. (2005). *District Laboratory Practice in Tropical Countries, Part 1, Second Edition*. Cambridge University Press, Cambridge, UK. 454 p.
- CHEESBROUGH M. (2006). *District Laboratory Practice in Tropical Countries, Part 2, Second Edition*. Cambridge University Press, Cambridge, UK. 434 p.
- DORNY P., BRANDT J. & GEERTS S. (2005). Detection and diagnosis. *In: WHO/FAO/OIE Guidelines for the Surveillance, Prevention and Control of Taeniosis/Cysticercosis*, Murrell K.D. ed. OIE, Paris, 45–55.
- EICHENBERGER R.M., LEWIS F., GABRIÉL S., DORNY P., TORGERSON P.R. & DEPLAZES P. (2013). Multi-test analysis and model-based estimation of the prevalence of *Taenia saginata* cysticercus infection in naturally infected dairy cows in the absence of a 'gold standard' reference test. *Int. J. Parasitol.*, **43** (10), 853–859.
- EICHENBERGER R.M., STEPHAN R. & DEPLAZES P. (2011). Increased sensitivity for the diagnosis of *Taenia saginata* cysticercus infection by additional heart examination compared to the EU-approved routine meat inspection. *Food Control*, **22**, 989–992.
- FLISSER A., CRAIG P.S. & ITO A. (2011). Cysticercosis and taeniosis *Taenia saginata*, *Taenia solium* and *Taenia saginata*. *In: Zoonoses. Biology, Clinical Practice, and Public Health Control*, Palmer S.R., Lord Soulsby E.J.L., Torgerson P.R. & Simpson D.I.H., eds. Oxford University Press, Oxford, UK, 625–642.
- GASSER R. & CHILTON N.B. (1995). Characterisation of taeniid cestode species by PCR-RFLP of ITS2 ribosomal DNA. *Acta Trop.*, **59**, 31–40.
- GONZALEZ L.M., MONTERO E., MORAKOTE N., PUENTE S., DIAZ DE TUESTA J.L., SERRA T. LOPEZ-VELEZ R., McMANUS D.P., HARRISON L.J., PARKHOUSE R.M. & GARATE T. (2004). Differential diagnosis of *Taenia saginata* and *Taenia saginata asiatica* taeniasis through PCR. *Diagn. Microbiol. Infect. Dis.*, **49**, 183–188.
- HERENDA D., CHAMBERS P.G., ETTRIQUEI A., SENEVIRATNA P. & DA SILVA T.J.P. (2000). Manual on meat inspection for developing countries. *FAO Animal Health and Production paper 119*. <http://www.fao.org/docrep/003/t0756e/T0756E00.htm>
- KHALIL L.F., JONES A. & BRAY R.A. (1994). *Keys to the Cestode Parasites of Vertebrates*. Wallingford, Oxon, UK: CAB International.
- LAWSON, J.R. & GEMMELL, M.A. (1990) Transmission of taeniid tapeworm eggs via blowflies to intermediate hosts. *Parasitology*, **100**: 143-146.
- LLOYD S. (2011). Other cestode infections. Hymenolepsis, diphyllbothriosis, coenurosis, and other adult and larval cestodes. *In: Zoonoses. Biology, Clinical Practice, and Public Health Control*, Palmer S.R., Lord Soulsby E.J.L., Torgerson P.R. & Simpson D.I.H., eds. Oxford University Press, Oxford, UK, 644–649.
- LLOYD S., MARTIN S.C., WALTERS T.M.H. & SOULSBY E.J.L. (1991). Use of sentinel lambs for early monitoring of the South Powys Hydatidosis Control Scheme: prevalence of *Echinococcus granulosus* and some other helminths. *Vet. Rec.*, **129**, 73–76.
- LOOS-FRANK B. (2000). An up-date of Verster's (1969) 'Taxonomic revision of the genus *Taenia*' (Cestoda) in table format. *Syst. Parasitol.*, **45**, 155–183.
- RODRIGUEZ S., WILKINS P. & DORNY P. (2012). Immunological and molecular diagnosis of cysticercosis. *Pathog. Glob. Health*, **106**, 286–298.

SCIUTTO E., MARTINEZ J.J., VILLALOBOS N.M., HERNANDEZ M., JOSE M.V., BELTRAN C., RODARTE F., FLORES I., BOBADILLA J.R., FRAGOSO G., PARKHOUSE M.E., HARRISON L.J. & DE ALUJA A.S. (1998). Limitations of current diagnostic procedures for the diagnosis of *Taenia solium* cysticercosis in rural pigs. *Vet. Parasitol.*, **79**, 299–313.

SOULSBY E.J.L. (1982). *Helminths, Arthropods and Protozoa of Domesticated Animals*, Seventh Edition. Balliere Tindall, London, UK, 809 p.

VAN KERCKHOVEN I., VANSTEENKISTE W., CLAES M., GEERTS S. & BRANDT J. (1998). Improved detection of circulating antigen in cattle infected with *Taenia saginata* metacestodes. *Vet. Parasitol.*, **76**, 269–274.

VERSTER A. (1969). A taxonomic revision of the genus *Taenia* Linnaeus 1758 s. str. *Onderstepoort J. Vet. Res.*, **37**, 3–58.

WALTHER M. & KOSKE J.K. (1980). *Taenia saginata* cysticercosis: a comparison of routine meat inspection and carcass dissection results in calves. *Vet. Rec.*, **106**, 401–402.

WANZALA W., ONYANGO-ABUJE J.A., KANG'ETHE E.K., ZESSIN K.H., KYULE N.M., BAUMANN M.P., OCHANDA H. & HARRISON L.J. (2003). Control of *Taenia saginata* by post-mortem examination of carcasses. *Afr. Health Sci.*, **3**, 68–76.

*

* *

NB: FIRST ADOPTED IN 1991. MOST RECENT UPDATES ADOPTED IN 2014.